
8 Chronic Effects of Acute, Low-Level Exposure to the Chemical Warfare Agent Sulfur Mustard*

Charles G. Hurst and William J. Smith

CONTENTS

- I. Introduction
 - II. Clinical Effects of Sulfur Mustard
 - A. Carcinogenesis
 - B. Chronic Pulmonary Disease
 - C. Chronic Eye Disease
 - D. Scarring, Pigmentation Changes, and Cancer of Epithelial Surfaces
 - E. Central Nervous System
 - F. Summary for Symptomatic Exposures
 - III. Acute Subclinical Exposure
 - A. Carcinogenesis
 - B. Radiation
 - IV. *In Vitro* Studies of Sulfur Mustard Toxicity
 - V. Dose Dependency of the Mustard Lesion
 - VI. Summary
- Acknowledgments
References

I. INTRODUCTION

Chemical warfare agents have been around for at least 4000 years and probably were originally used as poisons on individuals. The use of chemical weapons dates from at least 423 B.C. when allies of Sparta in the Peloponnesian War took an Athenian-held fort by directing smoke from lighted coals, sulfur, and pitch through a hollowed-out beam into the fort. Other conflicts during the succeeding centuries saw the use of smoke and flame. During the seventh century A.D., the Greeks invented “Greek fire.”

*The opinions or assertions contained herein are the private views of the authors, and are not to be construed as reflecting the view of the Department of the Army or the Department of Defense.

a combination probably of rosin, sulfur, pitch, naphtha, lime, and saltpeter that floated on water and was particularly effective in naval operations. During the fifteenth and sixteenth centuries, Venice employed unspecified poisons in hollow explosive mortar shells and sent poison chests to its enemy to poison wells, crops, and animals.¹⁻³ Finally, World War I and the Iran-Iraq War saw the advent of modern chemical warfare.

Mustard has been stockpiled in the arsenals of various countries since it was first used on July 12, 1917, when the Germans fired shells containing mustard at British troops entrenched near Ypres, Belgium. When a single agent was identified as the source of injury, it was estimated that mustard caused about 80% of the chemical casualties in World War I; other agents such as chlorine and phosgene caused the remaining 20%. The British had 180,983 chemical casualties; the injuries of 160,970 (88%) were caused solely by mustard. Of these casualties, 4,167 (2.6%) died. Of the 36,765 single-agent United States (U.S.) chemical casualties, the injuries of 27,711 (75%) were caused solely by mustard. Of the casualties who reached a medical treatment facility, 599 (2.2%) died. Just as disconcerting was the fact that mustard survivors required lengthy hospitalizations: the average length of stay was 42 days.⁴

Since the first use of mustard as a military weapon, there have been a number of isolated incidents in which it was reportedly used. In 1935, Italy probably used mustard against Abyssinia (now Ethiopia); Japan allegedly used mustard against the Chinese from 1937 to 1944; and Egypt was accused of using the agent against Yemen in the mid-1960s.

Chemical agents were not used during World War II. It is thought that Germany did not use mustard because Hitler had been a mustard victim during World War I and was loathe to use it.⁴

Sidell and Hurst have described the long-term effects produced from acute symptomatic clinical dose exposure to mustard, but less is known about the clinical effects from chronic, sometimes symptomatic, low-dose exposure.^{5,6} Acute is defined here as an exposure lasting less than 24 h. The term chronic refers to an exposure lasting for days, weeks, months, and even years. Clinical means producing a recognizable illness directly related to mustard exposure. Symptomatic means having either the acute or chronic clinical illness produced by sulfur mustard. Asymptomatic, of course, is without symptoms at all.

The argument can be made that acute, subclinical asymptomatic injury causing long-term effects does not exist. The pros and cons will be considered in this chapter, and parallels will be drawn. Certainly chronic, subclinical asymptomatic exposures do exist and there are parallels to other harmful situations. Obviously, workers in the manufacture of sulfur mustard, who were asymptomatic for part or all of their employment, fall into this category.

II. CLINICAL EFFECTS OF SULFUR MUSTARD

The organs most commonly affected by mustard are the skin, eyes, and airways: the organs which mustard contacts directly. After a substantial amount of mustard has been absorbed through the skin or inhaled, the hemopoietic system, gastrointestinal

tract, and CNS are also damaged. Mustard may also affect other organs but rarely do these produce clinical effects.⁴ After an asymptomatic latent period of hours, mustard causes erythema and blisters on the skin. This response ranges in severity from mild redness resembling sunburn to severe third-degree burns. Eye damage ranges from mild irritation-conjunctivitis, to corneal opacity, to perforation of the eye, and blindness. In the lung, the injury extends from mild upper respiratory signs to marked airway damage, bronchitis, and pneumonia. On rare occasions, acute laryngospasm can result in rapid death. Gastrointestinal effects vary from nausea and vomiting to severe hemorrhagic diarrhea. In the bone marrow, severe stem cell suppression can result in profound pancytopenia. In the CNS, at least in laboratory animals, seizures and death have been produced at high concentration exposures. The worst possible outcome from mustard exposure is death. However, mortality from mustard is uncommon. Less than 5% mortality from mustard gas was observed in allied troops in World War I.

Laboratory animal studies have shown that mustard is mutagenic and carcinogenic, and thus, it is not surprising that it is carcinogenic in man.⁷⁻⁹ Both Morgenstern et al. and Buscher and Green emphasize that chronic low-dose exposure over months to years in occupationally exposed workers leads to chronic bronchitis, bronchial asthma, hoarseness, aphonia, and hypersensitivity to smoke, dust, and fumes.^{10,11} Such individuals typically show persistent disability, with increased susceptibility to respiratory tract infections and evidence of bronchitis and bronchiectasis.¹⁰⁻¹²

All human studies dealing with chronic mustard disease processes are retrospective and fraught with the problems inherent in retrospective studies. These problems include bias in the sampling populations; lack of epidemiological controls from the effects of smoking, lifestyle, race, gender, age, or exposure to other chemicals; differential quality of available health care; and incorrect diagnosis.¹² These limitations make absolute interpretation of the studies difficult.

A. CARCINOGENESIS

Mustard is an alkylating agent similar to drugs that have been used in cancer chemotherapy, such as nitrogen mustards, Cytosan, and cis-platin. Since DNA is one of mustard's most sensitive targets, it is not surprising that carcinogenesis and radiomimetic effects are seen.⁵

Human data on the carcinogenicity of mustard are from (a) battlefield exposures, (b) accidents, and (c) workers in chemical factories. Both British and American studies have investigated the increased incidence of pulmonary carcinoma arising from World War I battlefield exposure. All are difficult to interpret, owing to the lack of controls for age, chronic pulmonary disease, cigarette smoking, and other factors that might affect the outcome.¹³⁻¹⁵

In contrast to battlefield exposures, studies of factory workers involved in the production of mustard have shown a definite link between prolonged exposure to low doses of mustard and cancer.¹² Several studies have provided evidence of an increased risk of respiratory tract cancers in factory workers.^{6,16-20} Easton et al. found a 45% increase in death due to lung cancer, a 170% increase in death from cancer of the larynx, and a 450% increase in death from cancer of the pharynx, compared with

expected deaths in the general population.¹⁷ The risks from cancer of the pharynx and lung were significantly related to the duration of employment at the factory.

B. CHRONIC PULMONARY DISEASE

Inhalation of mustard vapor primarily affects the laryngeal and tracheobronchial mucosa.¹² Evidence exists to suggest that mustard inhalation causes sustained respiratory difficulties even after the acute lesions have healed. Clinical follow-ups on 200 Iranian soldiers who were severely injured by mustard during the Iran-Iraq War indicate that about one third had experienced persistent respiratory effects 2 years or more after initial exposure. Reported problems included chronic bronchitis, asthma, rhinopharyngitis, tracheobronchitis, laryngitis, recurrent pneumonia, bronchiectasis, and, in some cases, severe, unrelenting tracheobronchial stenosis.²¹⁻²⁵

Of the British soldiers exposed to mustard in World War I, 12% were awarded disability compensation for respiratory disorders that were believed to be due to mustard exposures during combat.²⁶

Little contemporary information regarding the pathogenesis of the respiratory lesions is available, and few data from people or animals exposed to nonlethal concentrations of mustard vapor exist. Even fewer studies investigate the histopathology of the recovery process in animals exposed to mustard.⁹ However, two studies conducted during World War I suggest that low-level exposure or survivable exposures in dogs and rabbits may produce scar tissue following small ulcerations in the trachea and larynx, causing contractions of these areas.^{27,28} The more severe respiratory tract lesions described in animals exposed to mustard vapor appear to be quite similar in type and location to those described in humans.¹²

C. CHRONIC EYE DISEASE

Individuals who sustain acute ocular injury due to high-dose mustard exposure may experience difficulties even after the initial effects of the injury have subsided.²⁹⁻³² Recurrent or persistent corneal ulceration can occur after latent periods of 10 to 25 years. Chronic conjunctivitis and corneal clouding may accompany this delayed keratopathy.³¹⁻³² Anecdotal accounts suggest that low-dose exposure also causes increased sensitivity to later exposure to mustard, although the existence of increased sensitivity is difficult to substantiate with available scientific evidence.^{12, 33}

D. SCARRING, PIGMENTATION CHANGES, AND CANCER OF EPITHELIAL SURFACES

Skin cancer occurring at the site of old scar formation is an acknowledged biological phenomenon.^{34,35} Cutaneous cancers resulting from acute mustard exposure usually localize in scars, whereas those caused by chronic exposure can occur on any exposed site.³⁶

In a prospective study of delayed toxic effects from mustard exposure, Balali-Mood followed a group of Iranian soldiers exposed to mustard gas during the Iran-Iraq War.²⁴ After 2 years, 41% of the exposed victims were experiencing pigmentary disorders.

In the absence of melanocyte destruction, hyperpigmentation predominates. If melanocytes are locally destroyed, and inward migration from destroyed adnexal structures does not occur, depigmentation predominates.⁵

In its study of mustard and Lewisite effects, the Institute of Medicine concluded that, following mustard exposure:

- The evidence indicates a causal relation between acute, severe exposure to mustard agents and increased pigmentation and depigmentation in human skin.
- Acute and severe exposure can lead to chronic skin ulceration, scar formation, and the development of cutaneous cancer.
- Chronic exposure to minimally toxic and even subtoxic doses can lead to skin pigmentation abnormalities and cutaneous cancer.⁹

E. CENTRAL NERVOUS SYSTEM

Excitation of the CNS after mustard exposure, resulting in convulsions and followed by CNS depression, has been reported by the U.S. Army.³⁷ Convulsions and cardiac irregularities appear to occur only after extremely acute, high doses, which are probably attainable only in laboratory settings.^{12,38} Mustard casualties of the Iran-Iraq War did not display severe CNS or cardiac abnormalities.²¹

F. SUMMARY FOR SYMPTOMATIC EXPOSURES

The organs most commonly affected by mustard are the skin, eyes, and airways; the organs mustard comes in direct contact with. After a substantial amount of mustard has been absorbed through the skin or inhaled, the hemopoietic system, gastrointestinal tract, and CNS are also damaged. Mustard may also affect other organs, but rarely do these produce clinical effects.⁴ After an asymptomatic latent period of hours, mustard causes erythema and blisters on the skin. This ranges in severity from mild redness resembling sunburn, to severe third-degree burns. Eye damage ranges from mild irritation-conjunctivitis to corneal opacity, or even perforation of the eye and blindness. In the lung, the injury ranges from mild upper respiratory signs to marked airway damages, bronchitis, and pneumonia. On rare occasion, acute laryngospasm can result in rapid death. Gastrointestinal effects range from nausea and vomiting to severe hemorrhagic diarrhea. And in the bone marrow, severe stem cell suppression can result in profound pancytopenia. In the CNS, at least in laboratory animals, seizures and death have been produced at high concentration exposures. The worst outcome from all these organs systems, except for possibly the eye, is death. Death, however, is not the usual outcome from mustard exposure.

Studies of English and Japanese mustard factory workers establish repeated symptomatic exposures to mustard over a period of years as a causal factor in an increased incidence of airway cancer. The association between a single exposure to mustard and airway cancer is not as well established as the association between one-time mustard exposure and other chronic airway problems, such as chronic

bronchitis (based on World War I data). In some cases, the long-term damage was probably a continuation of the original insult resulting from insufficient therapy in the pre-antibiotic era. Morgenstern et al. give the following graphic description of symptoms and injuries incurred by some of the mustard factory workers.

Less widely known is the fact that many persons employed in the handling of mustard gas and exposed to small quantities of the vapor over a prolonged period of time may sustain damage to the respiratory mucosa, which may leave them partially or totally disabled. This statement is based on two and one-half years of observation in the medical department of an industrial plant where over 200 patients have been treated for both the acute symptoms and the residual effects of mustard gas exposure. The evolution of chronic mustard bronchitis may be traced as follows:

A young, white male previously engaged in farming or some other nonindustrial occupation with no history of any previous chronic lung disease goes to work on the mustard filling line. There is a varying concentration of mustard vapor in the air during a good part of the working day. After a period of time ranging anywhere from 3 weeks to 6 or 12 months he begins to show signs of definite irritation of the conjunctival and respiratory mucous membranes. He develops symptoms. He is given sick time off with his condition improving and returns to work. After a number of such episodes it becomes apparent that this man is not suitable for work in mustard and he is transferred out to another department free of toxic fumes.

After removal from mustard, his eyes and throat gradually heal. The conjunctivitis recedes and the vision returns to normal. The sore throat and hoarseness subside. The sense of taste returns, but the sense of smell may remain impaired. The appetite improves and he regains some of his lost weight with overall improvement. But he remains troubled by a persistent hacking cough, which come in paroxysms. It is most common in the morning but also occurs on lying down at night. It is often precipitated by physical exertion or when the man walks from the cold into a warm room or comes into contact with fumes of smoke. The cough is productive of anywhere from a teaspoon to a cupful of white or yellow mucoid or mucopurulent sputum, which may have a foul odor on occasion. There may be a troublesome wheezing and chest tightness most marked during damp weather. The patient seems to be more susceptible to respiratory infections than he was prior to exposure to mustard and the infections tend to last longer. Definite clinical bronchiectasis may develop as a result of repeated attacks of acute infectious bronchitis. He is hypersensitive to fumes and dust of any kind. He may develop dyspnea on slight or moderate exertion and therefore cannot perform any arduous labor.¹⁰

This description of chronic bronchitis developing in factory workers in the setting of World War II is both accurate and quite convincing.

Several eye diseases, such as chronic conjunctivitis, appear after an acute, usually severe, insult to the eye. In particular, delayed keratitis has appeared more than 25 years after the acute, severe lesion. Similarly, skin scarring, pigment changes, and even cancer have either followed the initial wound as a continuation of the process (scarring) or later appeared at the site of the lesion.

The production of nonairway cancer by mustard has been demonstrated in animals, but scant evidence exists to implicate mustard as a causative factor in nonairway cancer in humans.⁵

III. ACUTE SUBCLINICAL EXPOSURE

We are not convinced that acute, asymptomatic injuries that result in clinical disease truly exist. It is conceivable that certain synergistic situations can develop, such as would happen if co-factors or preexisting conditions (immunosuppression, genetic deficiency, or an additional chronic subclinical exposure) were triggered by some otherwise uneventful insult. This is certainly an unknown for mustard exposure at this time. The best that can be done is to draw analogies to other circumstances.

A. CARCINOGENESIS

Genotoxic substances usually have a direct effect on DNA and are occasionally effective after a single exposure. This helps to explain why they are frequently carcinogenic at subtoxic doses. These toxic compounds often act in a cumulative manner and synergistically with other DNA-reactive carcinogens. They usually produce neoplasms in more than one target organ and have a variable latency.³⁹

Strong promoters also possess weak intrinsic carcinogenicity. This experimental evidence comes from two test systems: (1) continued high-level administration of promoters such as croton oil to mouse skin or (2) oral administration of DDT or phenobarbital to rats. Both systems yield a small but definite crop of benign and malignant neoplasms in the absence of any obvious genotoxic carcinogen. An explanation is needed because promoters, by definition, do not have intrinsic properties of altering the genetic apparatus. One example has supplied an explanation that could apply to the others: when croton oil was applied to the skin of mice, it appeared to induce by itself a high incidence of papillomas and carcinomas.⁴⁰ Careful analysis revealed that the mice used were purchased from a supplier who housed the mice in creosoted cages. Thus, the mice had been exposed to genotoxic carcinogens in the creosote prior to the application of croton oil. Similarly, the weak carcinogenicity of DDT or phenobarbital may stem from prior exposure of the animals to small amounts of carcinogens, possibly mycotoxins or certain nitrosamines in the diet.⁴¹⁻⁴³

In mice, chemical enzyme-inducing substances have produced liver tumors, but were not found to be genotoxic using *in vitro* testing. Such chemicals have shown promoting activity in these systems.⁴⁴⁻⁴⁶ The liver of these animals appears to respond as if a DNA gene structure change has occurred. The role mycotoxins or nitrosamines play in the diet of these animals is open for speculation. Complex polychlorinated aliphatic and cyclic hydrocarbons also fall into the class of enhancing substances.^{47,48} They exhibit a nonlinearity in their dose-time response curves that differs from the genotoxic carcinogens.⁴⁹

The mechanism of promotion is subject to scientific research and considerable conjecture. The probability of multiple mechanisms is strong. A sequence of steps may be involved, leading to proliferation and differentiation. A parallel example might be the prostaglandin and cyclic nucleotide membrane effector systems. An early biochemical indicator of promotion is the induction of ornithine decarboxylase (ODC), and its presence has been used to discover new promoters.⁵⁰⁻⁵⁵ Increased levels of ODC appear to be associated with increased liver cell proliferation and might be related to the mechanisms of the promotion.^{56, 57}

Promoters and carcinogens involved in human cancer induction were first discovered through carefully conducted epidemiological studies, and then tested in animal models or in *in vitro* systems. Newer techniques use exfoliated cells or cells in culture to ascertain exposure to these genotoxic materials.⁵⁷⁻⁶⁰

Also, monoclonal antibody techniques may be able to trace carcinogen-macromolecular adducts in human tissues, verifying exposure to specific carcinogens.^{61,62} The gold standard will be to identify human hazards and carcinogens before exposure, “an ounce of prevention is worth a ton of cure.”⁶³⁻⁷⁰

B. RADIATION

Ultraviolet light is a form of nonionizing radiation exposure to man, while X-rays are an ionizing radiation. Both forms have the effect of damaging DNA similar to sulfur mustard, especially in rapidly dividing cells. It may never be possible to determine whether an individual can go through life without manifesting symptoms of acute sun damage: erythema, blisters, pigmentation, etc. or, at the very least, mild erythema. But, the chronicity of exposure producing the skin damage and cancers is undeniable. It is the acute symptomatic vs. chronic asymptomatic mix of exposures that will always remain a mystery.

Sulfur mustard has been called “radiomimetic” primarily because it appears to target DNA in rapidly dividing cells. There is a latent onset to its acute clinical symptoms very much paralleling those of ultraviolet sunlight damage, and a delayed onset to pulmonary cancers from chronic exposure. It is well established that the consequence of lifelong exposure to sunlight significantly enhances the development of skin cancers in fairer skinned individuals. The chronic damage to fair-skinned individuals is seen as premature aging, pigmentary changes, solar elastosis, solar keratosis, basal and squamous cell cancers, and malignant melanoma.^{71,72} The threshold for developing these lesions in man is multifactorial: heredity (skin type, individual’s ability to repair DNA), environment, and lifestyle. Thus, the cumulative dose responsible for these effects has great variability. Dark-skinned individuals may never experience any of the clinical entities mentioned above while individuals with the genetic disorder, Xeroderma pigmentosa, will experience marked acceleration of sun-related skin damage and cancers. This is caused by the genetic defect, which impairs the DNA repair produced by ultraviolet damage.⁷³

Likewise, it is well recognized that certain dyes, pigments, and drugs (i.e., tetracycline, psoralens, etc.) enhance sunlight and artificial light effects on skin. Existing genetic defects and specific drugs have not been identified that enhance the damage caused by sulfur mustard, but this possible synergism may be acute, subacute, or chronic.

Radiation-induced cancer is due to a nonlethal mutation of somatic cells. The latent period between irradiation and the development of cancer varies from 4 to 40 years, the average being 7 to 12 years. Even relatively low doses of X-rays increased the risk of cancer. Of school children epilated with 300 to 400 r of unfiltered 100-kV radiation, 1.6 percent had skin, thyroid, or parotid tumors 20 years later, while untreated control group showed only 0.2 percent such tumors. Apparently the mutated cells can survive 10 to 20 years before proliferating.⁷⁴

Evidence that various derivatives of tar and oil cause squamous cell carcinoma of the skin is both environmental and experimental. Experimental production of skin cancer in rodents with the various carcinogenic hydrocarbons has been well demonstrated. In fact, the effect of a chemical on animal skin is currently regarded as the best method of testing the carcinogenicity of the chemical.⁷⁵

IV. *IN VITRO* STUDIES OF SULFUR MUSTARD TOXICITY

Sulfur mustard is an alkylating agent that acts through cyclization of an ethylene group to form a highly reactive sulfonium electrophilic center. This reactive electrophile is capable of combining with any of the numerous nucleophilic sites present in macromolecules of cells. The products of these reactions are stable adducts which can modify the normal function of the target macromolecule. Since nucleophilic areas exist in peptides, proteins, RNA, DNA, and membrane components, extensive efforts have been underway to identify the most critical biomolecular reactions leading to mustard injury.

While the chemistry of mustard interactions with cellular components is well defined, the correlation of these interactions with injury has not been made. Over the past few decades, scientists have made major advances in understanding the cellular and biochemical consequences of exposure to mustard. While not the only target for alkylation by mustard, DNA is presumed to be an early reactant in the pathogenic cascades leading to the mustard lesion. Alkylation of nucleotides can result in apurinic site formation, disruption of normal DNA replication, activation of DNA repair pathways and eventually, to cytotoxic or mutagenic events. At high-exposure doses, such as those that lead to vesication *in vivo* or above 50 μM *in vitro*, the exposed cells sustain so much damage that they will die. The cells show activation of the nuclear enzyme poly(ADP-ribose) polymerase (PARP), disruption of cellular metabolism, loss of cell energetics, and total cellular breakdown. Many of these cells initially respond to the agent insult by activation of apoptotic death pathways, but in the absence of sufficient energy stores, quickly shift to a necrotic pattern of death. One could visualize this response as a cell suicide to eliminate the threat of long-term genotoxic sequelae such as mutation or cancer.

At low-dose exposure, the pattern could switch to one in which there is little to no immediate demonstration of injury but does set the stage for long-term consequences. As discussed previously, as a genotoxic agent, mustard can function in carcinogenesis in concert with, or playing the role of, promoter or inducer. The presence of nucleotide adducts or incorrect base replacement following DNA repair attempts coincidental with exposure to a second chemical insult could result in genesis of a cancer. But the question we wish to address is, "Can a low-dose, acute, asymptomatic exposure to mustard lead, by itself, to disease in later years?" Mustard is a mutagen and the mutation rate following *in vitro* exposure to cell culture systems has been studied. In the early 1970s, our laboratories reported that *in vitro* exposure of mouse lymphoma cells to submicromolar concentrations of mustard could increase the

reversion rate for asparagines independence. *In vivo*, using a host-mediated assay in mice, comparable reversion rates were seen with a single subcutaneous dose of 100 mg/kg.⁷⁶ In a concurrent study of chronic exposure over 37 weeks, mice were exposed in an inhalation chamber to 0.1 mg/m³ for 6 h per day, 5 days/week. No statistically significant increase in reversion rate was detected.⁷⁷ It is also interesting to note that the bifunctional alkylating agent sulfur mustard is less mutagenic than many monofunctional agents presumably because the cross-links formed by sulfur mustard lead to death of the affected cell.

Besides genotoxicity, sulfur mustard is known to affect other cellular parameters. Concentrations of mustard above 50 μM result in a marked reduction of cellular NAD⁺, a critical glycolytic cofactor, within 4 h of *in vitro* exposure to human keratinocytes.⁷⁸ However, no alterations in NAD⁺, ATP, or mitochondrial dehydrogenase activity are seen at 10-fold less concentrations in this model. One of the central cellular enzymes involved in NAD⁺ turnover is the nuclear enzyme PARP. As mentioned earlier, this enzyme undergoes a large and rapid activation in human epithelial cells exposed *in vitro* to vesicating equivalent doses (i.e., >50 μM). At concentrations of mustard below 10 μM , however, a completely different pattern of PARP response is seen.⁷⁹ It appears that even though significant DNA damage is detected at these low doses, the repair response is not as aggressive and the net metabolic disruption is transient.⁸⁰

As one studies the response to *in vitro* exposure to mustard in cell systems, there appears to be a threshold level above which death processes are initiated that are rapid and totally destructive to the cells. This appears to be in the range of 50–100 μM for most mammalian cells. If concentrations below 10 μM are studied, one can observe toxic processes occurring, but depending on the cell system employed, these are often reversible.

V. DOSE DEPENDENCY OF THE MUSTARD LESION

In 1946, Renshaw reviewed the understanding of the mechanisms of mustard injury to that point.⁸¹ He defined three dose ranges based on μg of mustard fixed per cm² of human skin. From 0.1–1.0 μg fixed/cm² the result was “mild erythema, occasional vesication with a histology that showed hyperemia and edema *without sufficient epidermal injury to cause death of more than occasional isolated basal cells*” (italics added). For 1.0–2.5 μg fixed/cm², the result was moderate injury with routine blister formation, and at doses >2.5 μg fixed/cm², the resulting injury was described as severe with central necrosis and circumferential vesication. Furthermore, he went on to state that minimal reversible injury was seen at 0.1 μg fixed/cm². One can say, therefore, at exposures resulting in less than 0.1 μg mustard fixed per cm² of skin, the outcome will be minimal clinical symptoms and fully reversible changes with no long-term effects. This exposure level is less than one-tenth that required for full demonstration of vesication and very close to what we refer to as a mild erythematous exposure.

Finally, for many years, sulfur mustard was used topically in the treatment of psoriasis in the form known as Russian Ointment (0.005% mustard-vaseline). This

was also known as Psoriasin. In the 1970s, Illig reviewed the clinical information from these studies and evaluated the potential skin carcinogenicity and off-gassing problems associated with cutaneous exposure to low-dose mustard.^{82,83} The following is a quote from his 1976 paper: “It is extremely improbable that the carcinogenic risk of the external S-mustard treatment is higher than that of a parenteral Methotrexate therapy, carried out at many clinics and in many cases over a period of many years, especially in the U.S.A.; it is rather to be expected that the carcinogenic risk in the case of external application of Psoriasin is also substantially lower over a longer period of time than in the Methotrexate treatment.”

VI. SUMMARY

Genotoxic agents have the potential of long-term consequences, especially when synergistically coupled with promoters, immunosuppression or genetic deficiencies. Our contention, that acute subclinical asymptomatic injury causing long-term effects does not exist, is based on the following:

1. Lack of reliable clinical cases
2. *In vitro* observations
3. Renshaw’s suggestion that, once the dose of applied mustard drops below that which yields observable, sustainable injury, no untoward consequences will become evident in the patient
4. More than 30 years’ experience with Russian Ointment

Based on our scientific and medical experience, we should never-say-never, but the probability of chronic illness developing from an acute asymptomatic exposure to sulfur mustard appears to be extremely low.

ACKNOWLEDGMENTS

The authors wish to acknowledge the assistance provided by the following: Patricia Little for editorial assistance and manuscript preparation, Cynthia Martinez and Bethany Toliver for collecting and organizing the references, and Clark Gross for reminding us of the papers on Russian Ointment.

REFERENCES

1. Joy, R.J.T., Historical aspects of medical defense against chemical warfare, in *Textbook of Military Medicine—Medical Aspects of Chemical and Biological Warfare*, Zajtchuk, R. and Bellamy, R.F., Eds., Office of The Surgeon General, Department of the Army, Washington, DC, 1997, chap. 3.
2. Smart, J.K., History of chemical and biological warfare: An American perspective, in *Textbook of Military Medicine—Medical Aspects of Chemical and Biological Warfare*, Zajtchuk, R. and Bellamy, R.F., Eds., Office of the Surgeon General, Department of the Army, Washington, DC, 1997, chap. 2.

3. *Medical Management of Chemical Casualties Handbook*, 3rd Ed., U.S. Army Medical Research Institute of Chemical Defense, Aberdeen Proving Ground, MD, 1999.
4. Sidell, F.R., Urbanetti, J.S., Smith, W.J., and Hurst, C.G., Vesicants, in *Textbook of Military Medicine—Medical Aspects of Chemical and Biological Warfare*, Zajtchuk, R. and Bellamy, R.F., Eds., Office of the Surgeon General, Department of the Army, Washington, DC, 1997, chap. 7.
5. Sidell, F.R. and Hurst, C.G., Long-term health effects of nerve agents and mustard, in *Textbook of Military Medicine—Medical Aspects of Chemical and Biological Warfare*, Zajtchuk, R. and Bellamy, R.F., Eds., Office of the Surgeon General, Department of the Army, Washington, DC, 1997, chap. 8.
6. Manning, K.P., Skegg, D.C.G., Stell, P.M., and Doll, R., Cancer of the larynx and other occupational hazards of mustard gas workers, *Clin. Otolaryngol.*, 6, 165, 1981.
7. Prokes, J., Svovoda, V., Hynie, I., Hroksova, M., and Keel, K., The influence of x-radiation and mustard gas on methionine-35-S incorporation in erythrocytes, *Neoplasma*, 5, 393, 1968.
8. Heston, W.E., Induction of pulmonary tumors in strain A mice with methyl-bis(beta-chloroethyl)amine hydrochloride, *J. Natl. Cancer Inst.*, 10, 125, 1949.
9. *Veterans at Risk: The Health Effects of Mustard Gas and Lewisite*, Pechura, C.M. and Rall, D.P., eds., The Institute of Medicine, Washington, DC, 1993.
10. Morgenstern, P., Koss, F.R., and Alexander, W.W., Residual mustard gas bronchitis: Effects of prolonged exposure to low concentrations of mustard gas, *Ann. Intern. Med.*, 26, 27, 1947.
11. Buscher, H. and Conway, N., *Green and Yellow Cross*, Cincinnati, OH, Kettering Laboratory of Applied Physiology, University of Cincinnati, OH, 1944.
12. Papirmeister, B., Feister, A.J., Robinson, S.I., and Ford, R.D., *Medical Defense against Mustard Gas: Toxic Mechanisms and Pharmacological Implications*, CRC Press, Boca Raton, FL, 1991.
13. Case, R.A.M. and Lea, A.J., Mustard gas poisoning, chronic bronchitis, and lung cancer: An investigation into the possibility that poisoning by mustard gas in the 1914–1918 war might be a factor in the production of neoplasia, *Br. J. Prev. Soc. Med.*, 9, 62, 1955.
14. Norman, J.R., Lung cancer mortality in World War I veterans with mustard gas injury: 1919–1965, *J. Natl. Cancer Inst.*, 54, 311, 1975.
15. Fletcher, C., Peto, R., Tinker, C., and Speizer, F.E., *The Natural History of Chronic Bronchitis and Emphysema*, Oxford University Press, Oxford, England, 1976.
16. Wada, S., Miyanishi, M., Nashimoto, Y., Kambe, S., and Miller, R.W., Mustard gas as a cause of respiratory neoplasia in man, *Lancet*, 1, 1161, 1968.
17. Easton, D.F., Peto, J., and Doll, R., Cancers of the respiratory tract in mustard gas workers, *Br. J. Ind. Med.*, 45, 652, 1988.
18. Minoue, R. and Shizushiri, S., Occupationally-related lung cancer—Cancer of the respiratory tract as sequentia from poison gas plants, *Jpn. J. Thorac. Dis.*, 18, 845, 1980.
19. Albro, P.W. and Fishbein, L., Gas chromatography of sulfur mustard and its analogs. *J. Chromatogr.*, 46, 202, 1970.
20. Yanagida, J., Hozawa, S., and Ishioka, S., Somatic mutation in peripheral lymphocytes of former workers at the Okunojima poison gas factory, *Jpn. J. Cancer Res.*, 79, 1276, 1988.
21. Willems, J.L., Clinical management of mustard gas casualties, *Ann. Med. Mil. Belg.* 3(Suppl), 1, 1989.
22. Urbanetti, J.S., Battlefield chemical inhalation injury, in *Pathophysiology and Treatment of Inhalation Injuries.*, Loke, J., Ed., Marcel Dekker, New York, 1988.

23. Balali-Mood, M., Clinical and laboratory findings in Iranian fighters with chemical gas poisoning, in *Proceedings of the 1st World Congress on New Compounds in Biological and Chemical Warfare: Toxicological Evaluation*, 21–23 May 1984, Heyndrickx B., Ed., State University of Ghent, Ghent, Belgium, 254, 1984.
24. Balali-Mood, M., First report of delayed toxic effects of yperite poisoning in Iranian fighters, in *Proceedings of the 2nd World Congress on New Compounds in Biological and Chemical Warfare: Toxicological Evaluation, Industrial Chemical Disasters, Civil Protection and Treatment*, 24–27 August 1986, Heyndrickx, B., Ed., University of Ghent, Ghent, Belgium, 489, 1986.
25. Freitag, L., Fizusian, N., Stamatis, G., and Greschuchna, D., The role of bronchoscopy in pulmonary complications due to mustard gas inhalation, *Chest*, 100, 1436, 1991.
26. Gilchrist, H.L., *A Comparative Study of World War Casualties from Gas and Other Weapons*, Government Printing Office, Washington, DC, 1928.
27. Warthin, A.S. and Weller, C.V., The lesions of the respiratory and gastrointestinal tract produced by mustard gas (dichloroethyl sulphide), *J. Clin. Lab. Med.*, 4, 229, 1919.
28. Winternitz, M.C., Anatomical changes in the respiratory tract initiated by irritating gases, *Mil. Surg.*, 44, 47, 1919.
29. Rimm, W.R. and Bahn, C.F., Vesicant injury to the eye, in *Proceedings of the Vesicant Workshop*, U.S. Army Medical Research Institute of Chemical Defense, Aberdeen Proving Ground, MD, 1987.
30. Hughes, W.F., Jr., Mustard gas injuries to the eyes, *Arch. Ophthalmol.*, 27, 582, 1942.
31. Blodi, F.C., Mustard gas keratopathy, *Int. Ophthalmol. Clin.*, 2, 1, 1971.
32. Duke-Elder, W.S. and MacFaul, P.A., Chemical injuries, in *System of Ophthalmology*, Duke-Elder, W.S. and MacFaul, P.A., eds., CV Mosby, St. Louis, MO, 1994.
33. Otto, C.E., *A Preliminary Report on the Ocular Action of Dichlorethyl Sulfide (Mustard Gas) in Man as Seen at Edgewood Arsenal, Edgewood, MD*, Edgewood Arsenal, Chemical Warfare Service, EAL 539, 1946.
34. Novick, M., Gard, D.H., Hardy, S.B., and Spira, M., Burn scar carcinoma: A review and analysis of 46 cases, *J. Trauma*, 17, 809, 1977.
35. Treves, N. and Pack, G.T., Development of cancer in burn scars: Analysis and report of 34 cases, *Surg. Gynecol. Obstet.*, 51, 749, 1930.
36. Inada, S., Hiragun, K., Seo, K., and Yamura, T., Multiple Bowen's disease observed in former workers of a poison gas factory in Japan with special reference to mustard gas exposure, *J. Dermatol.*, 5, 49, 1978.
37. U.S. Army, U.S. Navy, and U.S. Air Force, Vesicants (blister agents), Section I—Mustard and nitrogen mustard, in *NATO Handbook on the Medical Aspects of NBC Defensive Operations*, U.S. Army, U.S. Navy, U.S. Air Force, Washington, DC, AMedP-6, 1973.
38. Anslow, W.P. and Houch, C.R., Systemic pharmacology and pathology of sulfur and nitrogen mustards, in *Chemical Warfare Agents and Related Chemical Problems*, Office of Scientific Research and Development, Washington, DC, 1946.
39. Weisburger, J.H. and Williams, G.M., Bioassay of carcinogens: *In vitro* and *in vivo* tests, in *Chemical Carcinogens*, Searle, C. E., ed., ACS Monograph 182, Vol. 2, American Chemical Society, Washington DC, 1984, chap. 22.
40. Boutwell, R.K. and Bosch, D.K., The carcinogenicity of creosote oil: Its role in the induction of skin tumors in mice, *Cancer Res.*, 18, 1171, 1958.
41. Grice, H.C., Cleff, D.J., Coffin, D.E., Lo, M.T., Middleton, E.J., Sandi, E., Scott, P.M., Sen, N.P., Smith, B.L., and Withey, J.R., in *Carcinogens in Industry and the Environment*, Marcel-Dekker, New York, 1981, 439.

42. Walker, E.A., Castegnaro, M., Griciute, L., Börzönyi, M., and Davis, W., Eds., in *N-Nitroso Compounds: Analysis, Formation, and Occurrence*, IARC Scientific Publication No. 31, Lyon, France, 1980.
43. Silverman, J. and Adams, J.D., N-nitrosamines in laboratory animal feed and bedding, *Lab. Anim. Sci.*, 33, 161, 1983.
44. Berenblum, I., *Carcinogenesis as a Biological Problem*, Frontiers of Biology, North Holland, Amsterdam, 34, 1974.
45. Ford, J.O. and Pereira, M.A., Short-term *in vivo* initiation/promotion bioassay for hepatocarcinogens, *J. Environ. Pathol. Toxicol.*, 4, 39, 1980.
46. Williams, G.M., Phenotypic properties of preneoplastic rat liver lesions and applications to detection of carcinogens and tumor promoters, *Toxicol. Pathol.*, 10, 3, 1982.
47. Stott, W.T. and Watanabe, P.G., Differentiation of genetic vs. epigenetic mechanisms of toxicity and its application to risk assessment, *Drug Metab. Rev.*, 13, 353, 1982.
48. Stott, W.T., Reitz, R.H., Schumann, A.M., and Watanabe, P.G., Genetic and nongenetic events in neoplasia, *Food Cosmet. Toxicol.*, 19, 567, 1981.
49. Tennekes, H.A., Edler, L., and Kunz, H.W., Dose-response analysis of the enhancement of liver tumor formation in CF-1 mice by dieldrin, *Carcinogenesis*, 3, 941, 1982.
50. Hecker, E., Fusenig, N.E., Kunz, W., Marks, F., and Thielmann, H.W., Eds., *Cocarcinogenesis and Biological Effects of Tumor Promoters; Carcinogenesis—A Comprehensive Survey*, Raven Press, New York, 1982, 7.
51. Astrup, E.G. and Boutwell, R.K., Ornithine decarboxylase activity in chemically induced mouse skin papillomas, *Carcinogenesis*, 3, 303, 1982.
52. O'Brien, T.G., in *Polyamines in Biomedical Research*, Gaugas, J. M., Ed., Wiley Interscience, New York, 1980, 237.
53. Russell, K.H. and Haddox, M.K., Cyclic AMP-mediated induction of ornithine decarboxylase in normal and neoplastic growth, *Adv. Enzyme Regul.*, 17, 61, 1979.
54. Scalabrino, G. and Ferioli, M.E., Polyamines in mammalian tumors. Part I, *Adv. Cancer Res.*, 35, 151, 1981.
55. Fujiki, H., Suganuma, M., Nakayasu, M., Hoshino, H., Moore, R.E., and Sugimura, T., The third class of new tumor promoters, polyacetates (debromoaplysiatoxin and aplysiatoxin), can differentiate biological actions relevant to tumor promoters, *Gann*, 73, 495, 1982.
56. Izumi, K., Reddy, J.K., and Oyasu, R., Induction of hepatic ornithine decarboxylase by hypolipidemic drugs with hepatic peroxisome proliferative activity, *Carcinogenesis*, 2, 623, 1981.
57. Ide, F., Ishikawa, T., Takagi, M., Umemura, S., and Takayama, S., Unscheduled DNA synthesis in human oral mucosa treated with chemical carcinogens in short-term organ culture, *J. Natl. Cancer Inst.*, 69, 557, 1982.
58. Bruce, W.R. and Heddle, J.A., The mutagenic activity of 61 agents as determined by the micronucleus, salmonella, and sperm abnormality assays, *Can. J. Genet. Cytol.*, 21, 319, 1979.
59. Jenssen, D. and Ramel, C., The micronucleus test as part of a short-term mutagenicity test program for the prediction of carcinogenicity evaluated by 143 agents tested, *Mutat. Res.*, 75, 191, 1980.
60. Stich, H.F. and Rosin, M.P., Quantitating the synergistic effect of smoking and alcohol consumption with the micronucleus test on human buccal mucosa cells, *Int. J. Cancer*, 31, 305, 1983.
61. Perera, F.P., Poirier, M.C., Yuspa, S.H., Nakayama, J., Jaretski, A., Curnen, M.M., Knowles, D.M., and Weinstein, I.B., A pilot project in molecular cancer epidemiology: Determination of benzo [a] pyrene-DNA adducts in animal and human tissues by immunoassays, *Carcinogenesis*, 3, 1405, 1982.

62. Groopman, J.D., Haugen, A., Goodrich, G.R., Wogan, G.N., and Harris, C.C., Quantitation of aflatoxin B1-modified DNA using monoclonal antibodies, *Cancer Res.*, 42, 3120, 1982.
63. Gibson, J.L., Symposium: Peer review and scientific decision making, *Fundam. Appl. Toxicol.*, 2, 271, 1982.
64. Campbell, T.C., A decision tree approach to the regulation of food chemicals associated with irreversible toxicities, *Regul. Toxicol. Pharmacol.*, 1, 193, 1981.
65. Munro, I.C. and Krewski, D.R., Risk assessment and regulatory decision making, *Food Cosmet. Toxicol.*, 109, 549, 1981.
66. Starr, C. and Whipple, C., Risks of risk decisions, *Science*, 208, 1114, 1980.
67. Brown, S.M., The use of epidemiologic data in the assessment of cancer, *J. Environ. Pathol. Toxicol.*, 4, 573, 1980.
68. Vogt, T.M., Risk assessment and health hazard appraisal, *Ann. Rev. Public Health*, 2, 31, 1981.
69. Lave, L.B., Balancing economics and health in setting new standards, *Annu. Rev. Public Health*, 2, 183, 1981.
70. Scientific Committee Food Safety Council, Proposed system for food safety assessment, *Food Cosmet. Toxicol.*, 16, 1, 1978.
71. Urbach, F., *The Biologic Effects of Ultraviolet Radiation (with Emphasis on the Skin)*, Pergamon Press, Oxford, England, 1969.
72. Epstein, J.H. and Forbes, F.D., Ultraviolet carcinogenesis: Experimental, global and genetic aspects, in *Sunlight and Man*, Pathak, M.A., Harber, L.C., Leifick, M., and Kukita, A., Eds., University of Tokyo Press, Tokyo, Japan, 1974, 259.
73. Fornace, A.J., DNA single-strand breaks during repair of UV damage in human fibroblasts and abnormalities of repair in Xeroderma pigmentosum, *Proc. Nat. Acad. Sci., U.S.A.*, 73, 39, 1976.
74. Menon, I.A. and Haberman, H.F., Mechanisms of actions of melanins. *Br. J. Dermatol.*, 97, 109, 1977.
75. Poel, W.E., Skin as test site for the bioassay of carcinogens and carcinogen precursors, *Natl. Cancer Inst. Monogr.*, 10, 611, 1963.
76. Capizzi, R.L., Smith, W.J., Field, R.J., and Papirmeister, B., A host-mediated assay for chemical mutagens using the L5178Y/Asn(-) murine Leukemia, *Mutat. Res.*, 21, 6, 1973.
77. Rozmiarek, J., Capizzi, R.L., Papirmeister, B., Furman, W.H., and Smith, W.J., Mutagenic activity in somatic and germ cells following chronic inhalation of sulfur mustard, *Mutat. Res.*, 21, 13, 1973.
78. Smith, W.J., Gross, C.L., Chan, P., and Meier, H.L., The use of human epidermal keratinocytes in culture as a model for studying the biochemical mechanisms of sulfur mustard induced vesication, *Cell Biol. Toxicol.*, 6, 285, 1990.
79. Clark, O.E. and Smith, W.J., Activation of poly(ADP-ribose) polymerase by sulfur mustard in HeLa cell cultures, in *Proceedings of the 1993 Medical Defense Bioscience Review*, U.S. Army Medical Research Institute of Chemical Defense, Aberdeen Proving Ground, MD, DTIC Accession # A275667, 1, 199, 1993.
80. Smith, W.J., Toliver, B.S., Nealley, E.W., Guzman, J.J., and Gross, C.L., Effects of low dose sulfur mustard on growth and DNA damage in human cells in culture, *Toxicol. Sci.*, 54(1-S), 152, 2000.
81. Renshaw, B., Mechanisms in production of cutaneous injuries by sulfur and nitrogen mustard, in *Chemical Warfare Agents and Related Chemical Problems*, Bush, V., Ed., Office of Scientific Research and Development, National Defense Research Committee, Division 9, Parts 1-6, Washington DC, 1946, chap. 23.

82. Illig, L., The treatment of psoriasis vulgaris with S-mustard vasoline externally with special consideration to the possible carcinogenic risk (First continuation and conclusion): On the carcinogenicity of S-mustard in animal tests and in humans, *Z. Hautkrankh.*, 52, 1035, 1976.
83. Illig, L., Paul, E.L., Eyer, P., Weger, H., and Born, W., The treatment of psoriasis vulgaris with S-mustard-vaseline externally, taking especially into consideration the possible carcinogenic risk: III-Communication. Clinical and experimental studies on the extent of percutaneous and inhalative intake of S-mustard-vaseline, *Z. Hautkrankh.*, 54, 941, 1979.